



Natural Nanoparticles: A Particular Matter Inspired by Nature

Sharoon Griffin ^{1,2}, Muhammad Irfan Masood ^{1,3}, Muhammad Jawad Nasim ¹, Muhammad Sarfraz ¹, Azubuike Peter Ebokaiwe ⁴, Karl-Herbert Schäfer ³, Cornelia M. Keck ² and Claus Jacob ^{1,*}

- ¹ Division of Bioorganic Chemistry, School of Pharmacy, Saarland University, D-66123 Saarbruecken, Germany; sharoon.griffin@uni-saarland.de (S.G.); irfan_masood_79@yahoo.com (M.I.M.); jawad.nasim@uni-saarland.de (M.J.N.); s8musarf@stud.uni-saarland.de (M.S.)
- ² Institute of Pharmaceutics and Biopharmaceutics, Philipps University of Marburg, 35037 Marburg, Germany; cornelia.keck@pharmazie.uni-marburg.de
- ³ Department of Biotechnology, University of Applied Sciences Kaiserslautern, 66482 Zweibruecken, Germany; karl-herbert.schaefer@hs-kl.de
- ⁴ Department of Chemistry / Biochemistry and Molecular Biology, Federal University, Ndufu-Alike Ikwo, 482131 Ndufu-Alike, Nigeria; azubike.ebokaiwe@funai.edu.ng
- * Correspondence: c.jacob@mx.uni-saarland.de; Tel.: +49-681-302-3129

Received: 1 December 2017; Accepted: 22 December 2017; Published: 29 December 2017

Abstract: During the last couple of decades, the rapidly advancing field of nanotechnology has produced a wide palette of nanomaterials, most of which are considered as "synthetic" and, among the wider public, are often met with a certain suspicion. Despite the technological sophistication behind many of these materials, "nano" does not always equate with "artificial". Indeed, nature itself is an excellent nanotechnologist. It provides us with a range of fine particles, from inorganic ash, soot, sulfur and mineral particles found in the air or in wells, to sulfur and selenium nanoparticles produced by many bacteria and yeasts. These nanomaterials are entirely natural, and, not surprisingly, there is a growing interest in the development of natural nanoproducts, for instance in the emerging fields of phyto- and phyco-nanotechnology. This review will highlight some of the most recent—and sometimes unexpected—advances in this exciting and diverse field of research and development. Naturally occurring nanomaterials, artificially produced nanomaterials of natural products as well as naturally occurring or produced nanomaterials of natural products all show their own, particular chemical and physical properties, biological activities and promise for applications, especially in the fields of medicine, nutrition, cosmetics and agriculture. In the future, such natural nanoparticles will not only stimulate research and add a greener outlook to a traditionally high-tech field, they will also provide solutions—pardon—suspensions for a range of problems. Here, we may anticipate specific biogenic factories, valuable new materials based on waste, the effective removal of contaminants as part of nano-bioremediation, and the conversion of poorly soluble substances and materials to biologically available forms for practical uses.

Keywords: bioreduction; homogenization; microbes; nanoparticles; redox; selenium; sulfur; silver

1. Introduction

Today, nanotechnology and its diverse products are omnipresent and form an integral part of our products and lifestyle, from nanosilver in deodorants and nanoscopic particles with improved release properties in medicines all the way to "nanoimpregnations" of shower cabins, bath tubs and washing basins [1–3]. Whilst innovative materials containing particles with diameters in the one to one hundred nanometer range have emerged in many areas of our daily life, there has also been a feeling

that such materials are "not quite natural". Not surprisingly, therefore, the field of nano-toxicology more recently has attracted a particular interest—and there has also been mounting concern regarding a possible toxic impact on humans and contamination of the environment with nanomaterials [4].

This concern is certainly not entirely unjustified, as some dramatic examples, for instance asbestos (average diameter ranging from three to five micrometers) and other, air-bound fine particle matter, such as the PM_{2.5} fraction in exhaust gases, fumes and smoke illustrate [5–7]. Such critique, however, often ignores the fact that Nature itself is a skilled nanotechnologist, with numerous examples of common nanomaterials literally emanating from natural sources, such as volcanoes and mineral springs but also, in particular, from living organisms. Figure 1 provides a colorful reminder of such entirely natural sources of nanoscopic and microscopic particles. Indeed, life revolves around cells which themselves are microscopic in size (we do not account here for some rare and/or controversial nanobacteria) and metabolize molecules which are picoscopic, but also materials in between, which obviously are nanoscopic in their dimensions [8–10]. At the same time, nature also provides the inspiration and eventually also the ingredients—and even some of the methods—for natural nanomaterials.

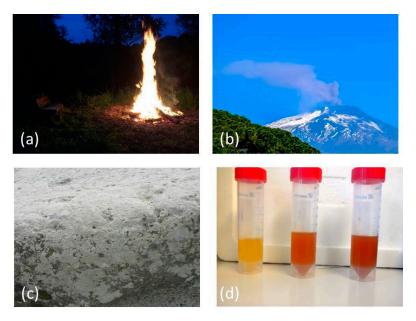


Figure 1. Nature itself is a skilled nanotechnologist. Microscopic and nanoscopic particles are formed, for instance, by combustion and are found: (**a**) near open fire; (**b**) as result of volcanic activity; (**c**) in form of precipitates; and (**d**) as bioreductively formed deposits of elements in certain bacteria. Photos provided by Marc Schäfer and Muhmmad Jawad Nasim.

Here, we will briefly consider the emerging field of natural nanoparticles. Before we start, we must, however, clarify what exactly is meant here. As any cunning linguist may have noted, the predicate "natural" may lead us into several directions. Indeed, there is a need for a distinction between nanomaterials which are, strictly speaking, natural, i.e., already present or formed in the environment without human intervention, and materials which are "nano" and also "bio". In the second case, the predicate "natural" equates with "biological", as in "natural products", which refers primarily to biological substances or materials [11–13]. Figure 2 illustrates this divide and provides a few selected examples of particularly interesting natural nanoparticles, nanoparticles of natural products and, eventually, naturally produced nanoparticles of natural products which we will discuss in more detail in the following sections. From the outset, we should emphasize that the field of natural nanotechnology is wide and diverse. Since we are unable to grasp it in its entirety, we will focus on a few highlights which we consider especially instructive, for instance in the fields of natural product-based nutrition, cosmetics, medicine or eco-friendly, "green" agriculture.

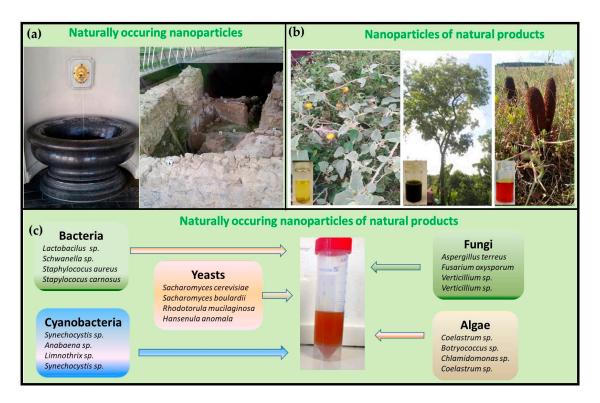


Figure 2. Examples of natural and biological materials which contain nanoscopic particles. (**a**) Naturally occurring nanoparticles of inorganic, elemental sulfur, for instance, are found at mineral wells rich in hydrogen sulfide, such as the Elisenbrunnen in Aachen. (**b**) In contrast, mechanically produced nanomaterials of natural products have been evaluated for medical and agricultural applications. (**c**) Eventually, there are also naturally produced nanomaterials of natural, biological products, such as nanoscopic particles of elemental selenium coated with microbial proteins which are formed by bioreductive or oxidative metabolism in bacteria and fungi.

2. Natural but Not Biological: The Free Flow of Inorganic Nanocomposites

Whilst hunting for nanoscopic materials in the environment, one soon realizes that a fair number of natural nanoparticles can be found outside the realm of life, for instance nanoscopic ash or soot particles as a result of volcanic activity, fires or other types of combustion. These particles are natural, yet usually not biological. Volcanic ash clouds contain a wide variety of polydisperse micro- and nanoparticles. These particles range from 100 to 200 nm in size and are chemically primarily composed of silicate and iron compounds. They are readily suspended in air and once inhaled may lead to serious respiratory disorders. Indeed, whilst particles of sizes in the lower micrometer range deposit in the upper respiratory tract, particles in the nanometer range penetrate deeply and deposit in tracheobronchial and alveolar regions where they can cause severe respiratory disorders [14]. "Carbon Nanotube" soot collected from the combustion of Texas Piñon Pine, for instance, contains multi-walled carbon nanotubes of 15 to 70 nm in size. These carbon-based objects readily become airborne and pose severe health hazards to animals and the human population [15].

Fire, however, is only one chemical process in the inorganic sphere which eventually may result in nanoscopic particles. Precipitation, oxidation and, to a lesser extent, reduction are also well suited to turn naturally occurring, inorganic materials into nanoparticles (Table 1).

Table 1. Selected examples of naturally occurring inorganic micro- and nanoparticles frequently found in our environment and associated with certain possible applications or biological implications. Please note that the practical applications mentioned usually rely on refined materials of particularly good particle quality and purity and not, or not yet, on the crude naturally occurring materials of similar constitution and composition.

Name	Chemical Formulae and Symbols	Natural Occurrence	Practical Implications (of Similar, Refined Materials)
Calcium Carbonate	CaCO ₃	natural surface water [16] 	industry, biotechnology, cancer therapy, drug delivery, plant nutrition and promotion of plant defense against pests [17–20]
Alumina	Al ₂ O ₃		desalination and defluorination of water [21–23]
Silicate	SiO ₄ ⁴⁻		drug carrier and catalytic applications [24,25]
Silica	SiO ₂	volcanic eruptions [26]	food additive, anti-caking agent, ultraviolet antireflection coating, cellular imaging and biomedical applications [27–30]
Bassanite (Calcium Sulfate)	CaSO ₄	sea water [31]	bone regeneration [32]
Iron Oxide	Fe ₃ O ₄	iceberg-hosted sediments [33]	medical diagnostics, controlled drug release, hyperthermia, biosensors, supercapacitor applications [34–37]
Manganese oxide	MnO ₂	umber [38]	imaging, remediation of contaminated soil and ground water, catalysis [39-41]
Sulfur	S	mineral wells [42]	medical applications, (antimicrobial, cytotoxic), fertilizers, fiber industry [43]
Soot (in the form of carbon)	С	atmospheric particulate matter	composite reinforcements, nano-reactors, chemical sensors, gas adsorbents, catalyst supports, templates, actuators, probes, nano-pipes [44,45]
Silver	Ag	aquatic environment [46,47]	antimicrobial properties, nano-functionalized plastics, paints, food containers, domestic appliances, textiles, medical products and cosmetics [46,48]
Gold	Au	ore deposits [49]	biosensorics, immunoassays, medical applications and laser phototherapy of tumors [50]
Platinum	Pt	automobile exhausts [51]	biomedical applications, nano-biomedicine, catalytic and thermal applications [52–54]

Amazingly, our drinking water, once considered under the microscope, is full of polydisperse nanoscopic as well as microscopic solid materials, of irregular shape and chemically based primarily on $CaCO_3$ and $CaSO_4$, often laced with other elements, such as iron oxides. Figure 3 provides an example of such deposits found near a mineral well in Aachen, Germany, and a microscopic view of the particles found in this water. Admittedly, such particles are of a rather poor quality and cannot be compared to the perfectly shaped, well-defined and homogeneous nanomaterials achieved by modern nanotechnological processes. Nonetheless, chemical, as well as physical processes, such as weathering, the slow precipitation of iron oxide particles but also dissolution and precipitation of carbonates under the influence of CO_2 and the intermediate formation of hydrocarbonate (HCO₃⁻), are able to generate such small-sized particle matter [55]. Abrasion, for instance, results in fine particle matter by scraping, cutting or grinding down larger lumps. There are numerous examples of natural nanomaterials formed this way, such as the $CaSO_4$ and silicate particles in spring water [16]. Indeed, such inorganic particles recently have inspired colleagues to synthesize a wide range of similar particles based on naturally occurring materials, such as—refined—nanoparticles of Fe₃O₄ and MnO₂ [56,57]. Concurrently, Nature, in cahoots with human activities, often unwillingly generates such "nanosized" particles from bulk materials, as the hot issue of microplastic in our oceans, fish and food highlights [58,59]. Nanoparticles generated by natural nanosizing are therefore not uncommon in our environment, and additional examples worth considering in earnest include the fine platinum particles released from millions of cars and their catalysts, as well as abrasion from tires, which are hardly "natural", yet slowly, but continuously, affect our environment and eventually also our health [60,61].

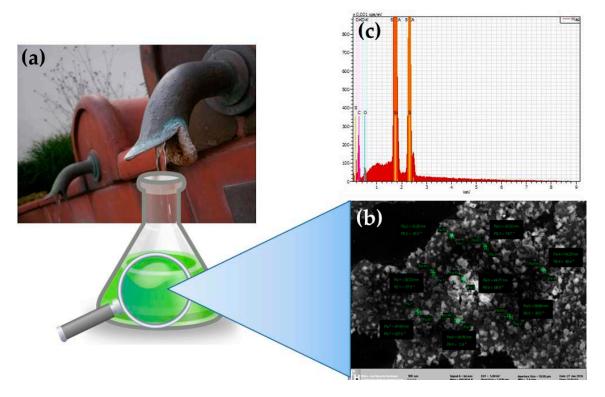


Figure 3. (a) The mineral wells in and near the town of Aachen in Germany are rich in sulfur, primarily in form of hydrogen sulfide (H₂S). Solid deposits of inorganic matter can therefore be found, for instance, at the Marktbrunnen in Burtscheid (image kindly provided by Roman Leontiev). (b) A microscopic investigation at 10,000-fold magnification reveals numerous microscopic and sub-microscopic particles and irregular agglomerates in this kind of water which (c) according to Energy Dispersion X-ray spectroscopy (EDX) consist of primarily of calcium salts and elemental sulfur [42].

Besides such "simple" physical and chemical events, there are similar, albeit more controlled particle generating processes, often based on spontaneous oxidation. One frequently observed example is the oxidation of hydrogen sulfide (H_2S) gas or hydrogen sulfide ions (HS^-) dissolved in waters of volcanic origin, such as volcanic lakes or mineral springs and wells which are common in many regions of the planet [62,63]. Here, the sulfide present in the water is oxidized by oxygen in the air to small particles of or containing elemental sulfur which can be found in the water itself, as in the Elisenbrunnen or Marktbrunnen in Aachen and, eventually, as part of larger sulfur deposits at or near the sulfide source. As before, Nature's inorganic chemistry is not the best nanotechnologist, and those particles are of a rather poor quality, polydisperse and also not entirely pure either. Still, they are already quite well defined—at least regarding chemical composition—and represent an interesting nanomaterial which is formed entirely naturally and can be obtained in considerable amounts and concentrations. From the perspective of resources, the flow of sulfur nanoparticles from volcanic sources is both, sustainable and virtually free of any costs. In the future, such natural sulfur nanoparticles from mineral wells may well be "harvested" or employed directly, for instance as a substitute for colloidal sulfur in agriculture, or may require some "maturation" in form of spontaneous or controlled oxidation of the hydrogen sulfide contained within the water. Nonetheless, simple medical applications, for instance on skin, also appear feasible. Indeed, many of the mineral wells particularly rich in sulfide, such as Bad Nenndorf or Bad Wiessee in Germany, offer bathing-rather than drinking—as part of therapy [64].

From a more scientific perspective, there are also still issues in the water which may need to be resolved. The active ingredient(s) in these wells, for instance, may well be simple sulfide (HS⁻), as traditionally assumed, but also inorganic polysulfides (HS_x⁻) or indeed elemental sulfur particles (S₈) [42]. Curiously, as these three classes of species exhibit their own, characteristic physical and chemical properties and reactivities, for instance in the context of the "cellular thiolstat" [65]. These Reactive Sulfur Species (RSS) are also easily converted into each other in the presence of oxidants (such as air), reductants (such as glutathione) or even spontaneously by mutual interactions in form of an extensive sulfur-centered "redox scrambling" [66]. Similarly, selenite (SeO₃²⁻) often occurs together with sulfur and, if reduced by H₂S or HS⁻, is able to form a wide range of elemental sulfur, selenium and mixed selenosulfur nanoparticles. As in the case of the sulfur particles in mineral wells, such selenosulfur species are interesting from a more applied, biological point of view. Selenosulfur compounds are well established and widely known, for instance, as active ingredients of certain anti-dandruff shampoos and even feature in movies such as "Evolution" [67,68].

Most of these nanocomposites generated more or less randomly in the wild by crude chemical processes are of an equally crude morphology and complex chemical composition. Good quality nanoparticles of elemental sulfur, selenium and tellurium can be produced under more controlled conditions in the laboratory employing a very similar "chemistry". The redox comproportionation of sulfide (HS⁻) and sulfite (SO₃²⁻), for instance, results in nanoscopic sulfur monodisperse particles of almost uniform size and round shape, and with an average diameter of around 150 nm. Similarly, the reduction of selenite (SeO₃²⁻) with the sulfur-containing amino acid L-cysteine yields spherical selenium nanoparticles with diameters in the range of 50 nm. Reduction of tellurite (TeO₃²⁻) with hydrazine (N₂H₄) even enables the generation of tellurium particles in the form of nanoscopic needles [69].

Inspiration for this kind of simple redox chemistry does not stop at the chalcogens. There are many other examples of spontaneous chemical (redox) transformations which eventually lead to small particles, such as the reduction of silver (Ag^+) or gold (Au^{3+}) cations to elemental silver or gold particles, respectively. In the following sections, we will therefore consider such reduction and oxidation reactions also in the context of other natural, biological agents.

3. Bioreductive Formation of Nanoparticles

Whilst volcanoes and mineral springs rely on simple chemical transformations to generate nanoparticles, living cells can recruit an entire arsenal of biotransformations to eventually produce such composites. Indeed, the living cell is mostly dealing with "nanotechnology", i.e., with objects of a nanoscopic size. Just to get a feeling for dimensions: A strand of DNA is 2.5 nm in diameter, a typical virus is around 100 nm wide and a typical bacterium is ten times bigger, i.e., in the range of $1-3 \mu$ m. Mammalian and plant cells are comparably large, occasionally reaching 50 μ m in diameter [70,71]. It is therefore not surprising that cells, and here microbial cells in particular, engage in some sort of "nanotechnology". Still, in the context of nanoparticles, there is one major caveat: Living cells normally do not prefer "the solid state", as any deposits formed intracellularly may stress and eventually kill them in a suicide-like process. The expulsion of such particles requires a more sophisticated machinery, effort and energy for excretion. Hence the formation of deposits inside cells is not that common—but also not entirely unknown either. When exposed to inorganic salts (e.g., containing S²⁻, SeO₃²⁻, Ag⁺ and Au³⁺), certain bacteria such as *Pseudomonas aeruginosa*, *Thiobacillus*, *Serratia*, and *Stenotrophomonas* species employ a reductive or oxidizing pathway of detoxification which eventually leads to the formation of elemental particles [72–77]. Such processes are rather well established and studied in the context of sulfur and selenium as well as silver nanoparticles, but also seem to apply to nanoparticles of gold and even platinum [78]. The particles produced by such biogenic factories are often of a surprisingly good quality, for instance small spherical shapes of an almost uniform size (Figure 4).

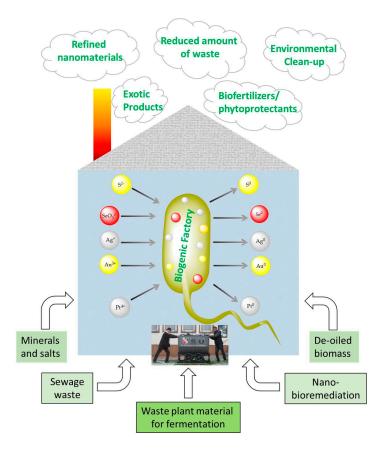


Figure 4. Schematic illustration of the biogenic factory which is able to turn biological substances, extracts, plants, algae and even waste biomass into amazing new products and nanomaterials. A particular interest resides on the added value resulting from the use and "up-cycling" of by-products and waste, such as de-oiled herbs, spent grains and coffee grounds, as these materials initially are not only food-grade but otherwise would go to waste and hence impact negatively on the environment. Photo taken at Hassel (Saar) and kindly provided by Elizabeth Jacob.

Not surprisingly, such biological processes may be exploited in practice to produce such particles of good quality and yield. This is the case, for instance, for selenium nanoparticles in dairy products produced by bacteria such as *Shewanella* sp. and *Lactobacillus* sp. Here, microorganisms which ferment the milk are also able to reduce selenite to elemental selenium for an additional "kick" [79,80]. Such processes are also of interest in the context of bioremediation and decontamination of soils enriched in certain toxic metals or semi-metals. Indeed, the removal of environmental contaminants (such as heavy metals, organic and inorganic pollutants) from contaminated sites using nanoparticles or nanomaterials formed by or in plants, fungi and bacteria with the assistance of nanotechnology, often referred to as nanobioremediation (NBR), is an emerging, environmentally friendly and economical alternative to traditional chemical methods [81,82]. Here, the three main strategies of modern bioremediation include the use of plants, microbes and isolated enzymes, for instance, laccase or nitrate reductase [83,84].

Whilst bioremediation is clearly an emerging topic related to microbially formed natural nanoparticles, it aims primarily at the removal of contaminants. Even so, there is also a more positive side to this approach. Here, the nanoparticles generated by such organisms are no longer seen as contaminants but actually as valuable nanomaterials of a more or less natural origin (Figure 4). Within this context, several of these bioreductively formed "natural" nanoparticles have been explored recently with sight on potential medical and agricultural applications [85–88].

It is possible, for instance, to recruit harmless microorganisms, such as *Saccharomyces cerevisiae* and *Staphylococcus carnosus* to generate fairly homogeneous selenium nanoparticles from SeO_3^{2-} with average diameters of 60 nm and 80 nm, respectively [89,90]. These particles can be harvested from the yeasts and bacteria after lysis of the cell. The authors of such studies have speculated about possible applications as food supplements and possibly as antimicrobial agents as some of these particles exhibit a certain antimicrobial activity [89–91]. In the field of agriculture, possible applications have even more facets, and a possible "hat trick" of simultaneously enriching the soil with selenium for fortified food products, of providing plants with elements for their natural defense systems and of eradicating plant pathogens seems feasible [89].

Within this context, one needs to emphasize that such naturally generated particles are not comparable to industrially generated materials. They are not "chemically pure", and usually also contain a "natural" coating of proteins whose composition is a reflection of the yeasts or bacteria they have been produced in. Hence the biological activity of such natural particles may stem from the bulk material of the particle itself, such as selenium, from other compounds trapped or contained within the particle, and also from the coating, which is often rich in proteins [89,92–94]. In such cases an extensive "intracellular diagnostics" is required to elucidate the exact target(s) and precise mode(s) of action [95].

Eventually, one may envisage an elegant process by which bacteria are grown on contaminated soils, and by remediating those soils produce well-defined nanoparticles which may be harvested and used in medicine, agriculture or other suitable applications. The resulting benefits of such an approach may be substantial—and are not far-fetched either, as relevant contaminants, such as heavy metals, often also represent the basis of particularly interesting particles.

As these strategies traditionally have focused primarily on the production of nanomaterials, the fate of the microorganisms involved usually has been of minor importance. Still, there may be some additional benefits, especially in the context of pathogenic fungi and bacteria. Several studies have demonstrated that the formation of nanosized materials by and inside pathogenic bacteria is an effective instrument to destroy those organisms. It has been noticed, for instance, that pathogenic strains of *Staphyllococcus aureus*, such as HEMSA and HEMSA 5M, reduce SeO_3^{2-} to elemental selenium when confronted with exceptionally high concentrations of this anion (around 2 mM) in an apparent attempt to deal with this exposure [96]. Eventually, this protective strategy fails and the deposits of selenium formed inside the bacteria kill these cells. This kind of "suicidal natural nanotechnology" is found among many bacteria and fungi, including pathogenic ones [89]. It partially explains in part the antimicrobial action often associated with SeO_3^{2-} and SeO_4^{2-} and other Reactive Selenium Species (RSeS), as well as with TeO₃²⁻ and TeO₄²⁻ [89]. Such activities may be specific for certain

organisms, endowing these agents and associated processes with certain "sensor/effector" properties. In the future, this kind of natural nanotechnology therefore may provide an interesting avenue to compromise, weaken, damage or perhaps even kill such pathogenic organisms [89].

4. Redox Chemistry with Natural Products

Just as many bacteria and fungi are able to produce nanoparticles of fairly good quality, this approach requires a certain effort in form of culturing and harvesting. It frequently also results in contamination with microbial biomolecules. Not surprisingly, alternative strategies have been developed which employ specific, isolated cellular components instead of whole cells to achieve the kind of-mostly bioreductive-chemistry which is usually required for the biological production of nanoparticles. As mentioned above, enzymes, such as laccase and nitrate reductase, are already employed in NBR, and similar avenues, based on isolated enzymes and simple natural reductants (or oxidants) have recently been explored as a means to generate nanoparticles [97–100]. Ascorbic acid, L-cysteine, reduced glutathione (GSH), flavonoids and a couple of other natural reducing agents are rather abundant in Nature and easy to obtain. Not surprisingly, these agents have been investigated already to produce nanoparticles of sulfur, selenium and silver, to name just a few [101-105]. Other redox active secondary metabolites, such as terpeniods (e.g., eugenol), flavonoids (e.g., luteolin and quercetin), sugars (e.g., glucose and sucrose) and certain amino acids (e.g., aspartate) have also been employed successfully to generate metal nanoparticles [101]. Besides simple plant metabolites, peptides have been considered, for example oligopeptides containing tryptophan residues. These peptides reduce metal ions to peptide-functionalized silver and gold nanoparticles [106]. Larger molecules, including redox active proteins, can also—chemically—produce nanoparticles, for instance particles of elemental platinum [107]. There are also reports that proteins from natural sources, such as whole cow milk, reduce metal cations, and generate, for instance, good quality nanoparticles of silver [108].

These few selected examples demarcate a particularly promising field of natural, biological nanotechnology, whereby isolated natural compounds, mixtures or even entire articles, such as whole milk, are used to produce nanomaterials. In practice, Nature provides a plethora of such reducing agents in form of compounds, peptides, proteins and enzymes. Indeed, certain microorganisms, plants and plant extracts are rich in antioxidants, with millimolar concentrations of ascorbic acid and thiols present therein [109–112]. As any lover of marmite will know, such extracts can be acquired rather easily, often as left-overs or by-products, such as yeast extracts from breweries. From an ecological and economical perspective, extracts are often superior to whole organisms and plants but also to isolated and extensively purified substances. Not surprisingly, therefore, such extracts are not only interesting from the prospect of being natural and fit for human consumption, but also since they are widely available and cheap [113,114]. Within this context, one rather noteworthy study has employed aqueous extracts of the fungus *Amylomyces rouxii* (strain KSU-09 isolated from the roots of *Phoenix dactylifera*) to generate silver nanoparticles [115].

In the context of plants, in particular, extracts are common and readily available. Not surprisingly, aqueous extracts of *Nelumbo nucifera* (root), *Embelia ribes* (seed), *Rosmarinus officinalis*, *Ocimum basilicum*, *Petroselinum crispum* (leaf and root), *Citrus limon* (peel), *Vitis vinifera* (peel), *Cucumis sativus* (peel), *Mimusops elengi* Linn. (leaf), *Acalypha indica* (leaf), *Zingiber officinalis* and *Capsicum frutescens*, among others, are frequently used to produce nanoparticles of Ag, Au, Fe₃O₄ and ZnO [116–124]. Indeed, this emerging field of "phyto-nanotechnology" provides numerous advantages (Figure 5). The materials employed, such as extracts of herbs, are often available as cheap by-products, yet still rich in active ingredients, and therefore of value for further processing. One should also remember that some of these plant products are "food grade", and hence entail possible applications in nutrition and cosmetics [125]. Together with other "readily available" biomass, such as microalgae, such materials are well suited for the controlled synthesis of good quality nanoparticles [126]. Indeed, the emerging field of "phyco-nanotechnology" relies explicitly on algae for bio-nanomanufacture as these organisms

are not only highly interesting from a scientific point of view, but also readily available, easy to culture and environmentally friendly to use [127].

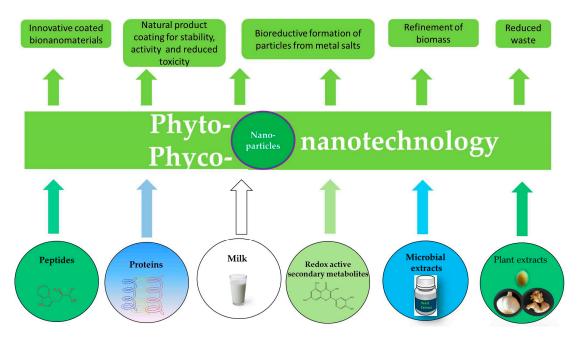


Figure 5. The emerging field of phyto-nanotechnology employs isolated biological components and substances to form, modify or coat nanoparticles. These particles often exhibit interesting properties, such as pronounced biological activity, and may therefore be employed in medicine or agriculture. Phyto-nanotechnology also offers new and innovative uses for plant materials and biomass, which otherwise may have been wasted. Here, the field of phyco-nanotechnology, which is centered around algae, for many biological, manufacturing, ecological and economical reasons today represents a particularly interesting area of research and development.

Besides these more obvious applications of extracts and by-products in phyto-nanotechnology, one should also briefly mention two additional applications in this field. One is the use of such "waste" biomass as feedstock for bacteria and fungi able to generate nanoparticles in vivo [101,128]. In this case, the biomass is not used directly as reducing material, as above, but rather indirectly—and probably more extensively—to promote the growth of suitable bacteria able to perform this kind of bioreduction. The second application concerns the coating of nanoparticles. As mentioned above, nanomaterials produced by bacteria are often coated with proteins, and this coating may endow such particles with improved stability, further features and especially also additional biological activities. It is therefore not surprising that natural substances, such as extracts of Darjeeling tea, have been investigated as coatings for silver nanoparticles to provide stability against agglomeration and also to reduce toxicity [129].

In general, these materials—literally—provide a fertile ground for future research and development, especially in the context of turning biomass waste into (nanomaterial) value (Figure 5).

5. Milling Vanilla

Thus far, natural products have been employed primarily to generate, cover or coat nanoparticles. This raises the question if such biological materials themselves may not be converted into nanoparticles. Similar to naturally occurring abrasion mentioned earlier, methods such as grinding, milling and (high pressure) homogenization provide a wide and colorful arsenal of methods able to "mill down" almost any material, including chemical elements in their solid state, sparingly soluble food supplements and medications, and, actually, also plant parts and even entire trees [130]. The resulting particles of such

natural products are of a unique nature, as they are still natural products, yet have been transformed into an unusual, unnatural size and shape.

It is therefore hardly surprising that many natural products have been nanosized (or "nanonized") during the last couple of years. Antioxidants such as rutin, for instance, have been turned into so-called "nanocrystals" using an eloquent technique which involves wet bead milling (WBM) and high-pressure homogenization (HPH) [131,132]. Here, nanotechnology can be used to produce nanoparticles with a dramatically improved solubility, excellent release kinetics and hence a good bioavailability and biological activity. This approach is particularly attractive in the field of—often sparingly soluble—antioxidants and plant products rich in such antioxidants, i.e., substances and materials which originally have poor release kinetics on the lipid/aqueous surface of the skin but thanks to the new technology can nowadays be used easily, for instance in cosmetics.

The basic physical principles behind this approach of nanosizing natural products are illustrated in Figure 6. Indeed, the principle of nanosizing coarse materials to improve their biological activity is very simple and is mainly based on the Noyes–Whitney equation, one of the major equations in biopharmacy (Equation (1)).

$$\frac{dc}{dt} = D \cdot A \cdot \frac{(c_s - c_0)}{h} \tag{1}$$

where dc/dt is the dissolution rate, *D* the diffusion coefficient, *A* the total surface area of the particles, c_s the saturation solubility of the active ingredient, c_0 the concentration of dissolved active ingredient in the solvent and *h* the diffusional distance.

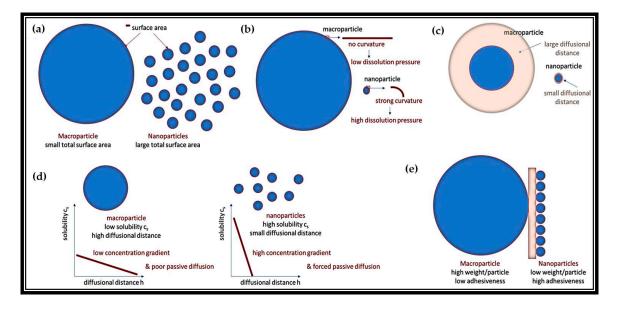


Figure 6. Properties of nanoparticles: (**a**) increased rate of dissolution; (**b**) enhanced saturation solubility; (**c**) decreased diffusional distance; (**d**) higher concentration gradient; and (**e**) improved adhesiveness.

When nanosizing coarse material the rate of dissolution dc/dt increases, because the total surface area of the particles involved increases (Figure 6a). In this case, the saturation solubility increases due to a higher dissolution pressure, which is explained by a higher curvature of the particles (Kelvin equation, Figure 6b) and the diffusional distance is also decreased (Prantl equation, Figure 6c). Eventually, nanosizing leads to a significant increase in the overall velocity of dissolution, which is especially interesting if active ingredients dissolve slowly or are even poorly soluble in water. Furthermore, nanosizing improves the bioactivity of poorly soluble active ingredients. As a result of the increase in solubility the concentration gradient, when compared to larger sized materials, is increased [133]. If the active ingredient is taken up by the body (or the plant) via passive diffusion, the concentration gradient is the driving force for uptake or permeation. Hence the higher the concentration gradient, the faster and more efficient the uptake will be (Figure 6d). Eventually, the smaller the size of a particle, the faster it will dissolve and the higher its bioactivity will be [134].

There are also additional benefits. Nanosized materials possess a much higher adhesiveness to surfaces than coarse materials. This is due to the much larger surface to volume ratio of nanoparticles, which translates into considerably more attaching points per volume and therefore to less forces needed to stick to a surface (Figure 6e). Hence, after administration or application, nanoparticles tend to adhere much tighter and longer to surfaces, the time to dissolve and to penetrate is prolonged, therefore further increasing the bioactivity of active ingredients. Due to these superior features and the ease of production, nanosizing, i.e., the production of nanocrystals, has become a major formulation principle in pharmaceutics to improve the bioactivity of pharmaceutically active ingredients [135–137].

In the field of natural products, nanosizing has opened up a promising avenue to augment further their potential. It is possible, for instance, to convert simple, intrinsically "insoluble" materials, such as elemental sulfur, selenium and tellurium—and, of course, many of the other solid elements of the Periodic Table—into nanosuspenions with interesting biological activities [69]. In the case of the three chalcogens, a pronounced biological activity, for instance against *Steinernema feltiae*, *Escherichia coli* and *Saccaromyces cerevisiae* has been observed which compares well with the one of the corresponding elemental particles obtained by redox chemistry or bioreduction in *S. carnosus* [89].

Nanosizing chemically pure substances or mixtures is comparably straight forward, yet the matter literally becomes more complicated once natural samples such as barks, shells, seeds or even dried fruits, roots or entire plants are milled and homogenized. Here, specific techniques need to be developed and applied. It may appear straight forward to mill down a shell of a walnut or some commercially available grape seed flour, even so our experience tells us that nanosizing a dried tomato plant or spent coffee ground is considerably more challenging. Specific protocols with several steps may be required, from drying and freeze-drying to defatting and pre-milling. Still, some plants, such as the Maltese mushroom *Cynomorium coccinem* L., a parasitic plant devoid of any chlorophyll, can be obtained, freeze-dried, milled and homogenized to fairly stable, low polydispersity and uniform particles with average diameters of around 400 nm without encountering any unsurmountable problems [138]. Some of these milled down plant materials have been evaluated already as potential food supplements and even as natural medicines and antimicrobial agents [130]. In the medium term, nanoparticles of natural products may be employed in the fields of nutrition, medicine, cosmetics or, in the case of large scale manufacture, in "green" agriculture (Figure 7).

The activities observed for those nanosized materials are often promising, yet there is a fine balance of arguments which needs to be considered. On a positive note, nanosizing an herb or medical plant is comparably straightforward and considerably easier than extraction, purification and formulation of the active ingredient(s) contained therein. It also produces no or little waste. Furthermore, the nanoparticles essentially are still "natural", at least as far as their chemical composition is concerned, contain all the ingredients of the plant, have not undergone any extensive modifications and, notably, have not been treated with any organic solvents. Ideally, they even represent a natural slow release system of bioavailable and biologically active ingredients. In the case of HPH, such materials initially are also sterile and, as more recent studies have confirmed, can also be lyophilized and resuspended without loss of physical properties or activity once a simple stabilizer such as mannitol is added [139–141].

Eventually, some caution is required as such materials are intrinsically ill defined chemically, often contain fibrous materials, are prone to fouling if contaminated with microorganisms and also require certain stabilizers so not to aggregate in nanosuspension or as a result of freeze-drying. In analogy to the 1980s German pop band "Milli Vanilli", milling vanilla is clearly exciting, fancy, hot and full of potential, yet some care must be taken and there is still considerable need for further investigation and improvement, especially once the power fails and the chips are down [142].

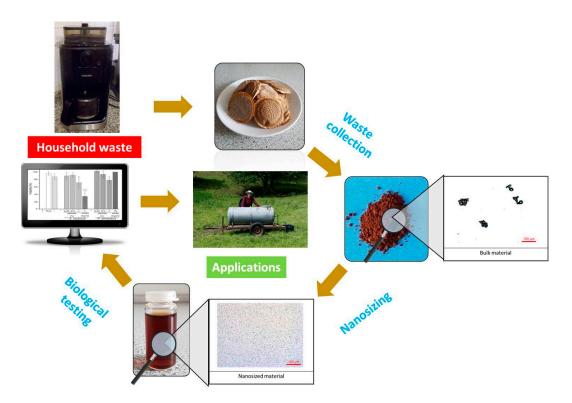


Figure 7. A schematic overview of theutilization of nanosizing techniques for turning waste into value.

6. Conclusions

The previous sections have highlighted just a few selected recent developments at the interface of nanotechnology and natural products research. We have seen that nature itself is well suited to produce a repertoire of nanomaterials by processes such as combustion, abrasion, precipitation and oxidation, and, if the biosphere is included, by bioreduction and related processes. Some of the natural nanoparticles obtained in this way may be useful in medicine, agriculture or other fields of technology and engineering. Not surprisingly, this has stimulated research into these native materials and processes, and has also inspired strategies to generate biological and biologically active materials using similar materials and methods. It is now time to take stock and to anticipate some of the most exciting developments which the next couple of years may bring (Figure 8).

First, it seems plausible that some freely flowing inorganic substances, such as H₂S, which hitherto have "only" been used in spa towns to pickle and macerate affluent pensioners or have been wasted entirely, may be reconsidered as valuable precursors of fine chemicals, including certain nanoparticles. Here, some "hat tricks" may also be feasible, such as the reaction of H₂S from mineral springs with fumes rich in SO₂ as part of an elegant sulfur redox comproportionation, or a reaction of H₂S with NO₂. Such "waste chemistry" may not only be employed to produce the desired nanoparticles—in this case of sulfur, but also to detoxify two individual environmental hazards simultaneously [42]. Those ideas are still speculative, however, early studies into this direction are marred by issues, such as adequate concentrations and how to bring the hazards—literally—under one roof. It seems to us that crucial but manageable research is required to define the correct ingredients and conditions for such manufacture and large-scale production of particles of sulfur and related elements, such as selenium.

Secondly, natural nanotechnology employing organisms such as yeasts and certain harmless bacteria, but also isolated enzymes, may in future be recruited to generate a variety of particular particle matter, starting with selenium and embracing large parts of the Periodic Table, but also other inorganic materials, such as insoluble metal oxides. Indeed, it seems today that a wide palette of insoluble matter may be generated inside bacteria. Whilst traditionally harvesting of such particles has involved lysis of cells, some organisms also release their particles into the supernatant, as has been demonstrated

for resveratrol-conjugated gold nanoparticles produced by the *Delftia* sp. strain KCM-006 [143]. Such in vivo generation of nanoparticles may provide further impetus for bioremediation and inspire new avenues to tackle some pathogenic organisms unable to release their particles with undesired intracellular deposits.

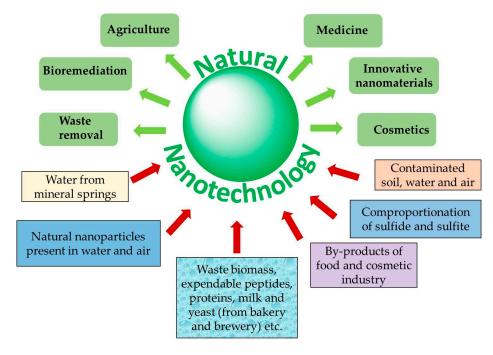


Figure 8. Natural nanotechnology with its various inorganic and biological aspects provides a wide range of opportunities and applications, not only in medicine and cosmetics, but also in less obvious areas such as agriculture and waste removal.

Thirdly, the use of certain plant products, such as de-oiled herbs, as reagents in nano-manufacture may be considered more widely, especially in the context of waste management and "up-cycling". Here, plant waste from harvests or processes such as baking or brewing may provide an interesting alternative to pure chemicals, as they are natural and readily available in large quantities and at low cost. As mentioned already, these materials may either be employed directly as reducing agents or coatings or, more indirectly, as feedstock for bacteria able to generate nanoparticles via a bioreductive avenue. From an ecological perspective, it would be especially intriguing to employ nanosized waste as feedstock for bacteria naturally producing natural nanoparticles.

Fourthly, milling and homogenization of plants may unlock a whole treasure chest of new products, as it can be employed to render hitherto insoluble materials into nanosuspensions with interesting release properties. Many of these products originate in agriculture and, once processed, may be used there as well, hence providing the basis for interesting production and application cycles. This field is still in its early stages of research and development and most certainly will lead to many obstacles and pitfalls, but also to some truly innovative ideas, methods and products.

Eventually, we are likely to witness a rapidly growing interest in various fields of bio-nanotechnology, such as phyto- and phyco-nanotechnology, not only with sight on product development, but also in many areas of basic research which accompany such developments. In the longer run, it may even be possible to explore some of these leads to generate nanoparticles of natural products, such as active ingredients of plants, using natural processes, including bacterial or fungal fermentation, in vitro bioreduction or abrasion. Here, the two meanings of natural, i.e., in form of the material or as part of the method, may eventually meet and merge. Most of this is obviously still speculative today, and time will tell which of these leads are green, fruitful inspirations and which are more the kind of red herring which will stay in the fishbowl of the laboratory.

Acknowledgments: The authors would like to acknowledge their respective Universities for financial support: The University of Saarland, University of Applied Sciences Kaiserslautern, Federal University of Ndufu-Alike Ikwo and the Philipps University of Marburg. The authors would also like to acknowledge the financial support provided by Higher Education Commission of Pakistan and TWAS-DFG. Special thanks go to the members of the "Academiacs International" network for their helpful discussions and advice.

Author Contributions: Sharoon Griffin, Muhammad Irfan Masood, Muhammad Jawad Nasim, Muhammad Sarfraz, Azubuike Peter Ebokaiwe, Karl-Herbert Schäfer, Cornelia M. Keck and Claus Jacob wrote the manuscript.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Kettler, K.; Krystek, P.; Giannakou, C.; Hendriks, A.J.; de Jong, W.H. Exploring the effect of silver nanoparticle size and medium composition on uptake into pulmonary epithelial 16HBE14o-cells. *J. Nanopart. Res.* 2016, 18, 1–11. [CrossRef] [PubMed]
- 2. Moss, D.M.; Siccardi, M. Optimizing nanomedicine pharmacokinetics using physiologically based pharmacokinetics modelling. *Br. J. Pharmacol.* **2014**, *171*, 3963–3979. [CrossRef] [PubMed]
- 3. Vincent, B.B.; Loeve, S. Metaphors in nanomedicine: The case of targeted drug delivery. *NanoEthics* **2014**, *8*, 1–17. [CrossRef]
- 4. Roy, D.N.; Goswami, R.; Pal, A. Nanomaterial and toxicity: What can proteomics tell us about the nanotoxicology? *Xenobiotica* **2017**, 47, 632–643.
- Brunner, T.J.; Wick, P.; Manser, P.; Spohn, P.; Grass, R.N.; Limbach, L.K.; Bruinink, A.; Stark, W.J. In vitro cytotoxicity of oxide nanoparticles: Comparison to asbestos, silica, and the effect of particle solubility. *Environ. Sci. Technol.* 2006, 40, 4374–4381. [CrossRef] [PubMed]
- 6. Cassee, F.R.; Heroux, M.E.; Gerlofs-Nijland, M.E.; Kelly, F.J. Particulate matter beyond mass: Recent health evidence on the role of fractions, chemical constituents and sources of emission. *Inhal. Toxicol.* **2013**, *25*, 802–812. [CrossRef] [PubMed]
- Kukkonen, J.; Bozó, L.; Palmgren, F.; Sokhi, R.S. Particulate matter in urban air. In *Air Quality in Cities:* Saturn Eurotrac-2 Subproject Final Report; Moussiopoulos, N., Ed.; Springer: Berlin/Heidelberg, Germany, 2003; pp. 91–120.
- 8. Urbano, P.; Urbano, F. Nanobacteria: Facts or fancies? *PLoS Pathog.* 2007, *3*, 567–570. [CrossRef] [PubMed]
- Kajander, E.O.; Ciftcioglu, N.; Miller-Hjelle, M.A.; Hjelle, J.T. Nanobacteria: Controversial pathogens in nephrolithiasis and polycystic kidney disease. *Curr. Opin. Nephrol. Hypertens.* 2001, 10, 445–452. [CrossRef] [PubMed]
- Ciftcioglu, N.; Mckay, D.S.; Mathew, G.; Kajander, E.O. Nanobacteria: Fact or fiction? Characteristics, detection, and medical importance of novel self-replicating, calcifying nanoparticles. *J. Investig. Med.* 2006, 54, 385–394. [CrossRef] [PubMed]
- 11. Pasula, R.R.; Lim, S. Engineering nanoparticle synthesis using microbial factories. In *Engineering Biology*; Institution of Engineering and Technology: Stevenage, UK, 2017; Volume 1, pp. 12–17.
- 12. Patel, V.; Berthold, D.; Puranik, P.; Gantar, M. Screening of cyanobacteria and microalgae for their ability to synthesize silver nanoparticles with antibacterial activity. *Biotechnol. Rep.* **2015**, *5*, 112–119. [CrossRef] [PubMed]
- Mukherjee, P.; Ahmad, A.; Mandal, D.; Senapati, S.; Sainkar, S.R.; Khan, M.I.; Ramani, R.; Parischa, R.; Ajayakumar, P.V.; Alam, M.; et al. Bioreduction of AuCl₄⁻ ions by the fungus, *Verticillium* sp. and surface trapping of the gold nanoparticles formed. *Angew. Chem. Int. Ed.* 2001, 40, 3585–3588. [CrossRef]
- 14. Lahde, A.; Gudmundsdottir, S.S.; Joutsensaari, J.; Tapper, U.; Ruusunen, J.; Ihalainen, M.; Karhunen, T.; Torvela, T.; Jokiniemi, J.; Jarvinen, K.; et al. In vitro evaluation of pulmonary deposition of airborne volcanic ash. *Atmos. Environ.* **2013**, *70*, 18–27. [CrossRef]
- 15. Murr, L.E.; Guerrero, P.A. Carbon nanotubes in wood soot. Atmos. Sci. Lett. 2006, 7, 93–95. [CrossRef]
- 16. Wu, C.Y.; Martel, J.; Wong, T.Y.; Young, D.; Liu, C.C.; Lin, C.W.; Young, J.D. Formation and characteristics of biomimetic mineralo-organic particles in natural surface water. *Sci. Rep.* **2016**, *6*. [CrossRef] [PubMed]
- 17. Boyjoo, Y.; Pareek, V.K.; Liu, J. Synthesis of micro and nano-sized calcium carbonate particles and their applications. *J. Mater. Chem. A* **2014**, *2*, 14270–14288. [CrossRef]

- Koo, A.N.; Min, K.H.; Lee, H.J.; Jegal, J.H.; Lee, J.W.; Lee, S.C. Calcium carbonate mineralized nanoparticles as an intracellular transporter of cytochromec for cancer therapy. *Chem. Asian J.* 2015, *10*, 2380–2387. [CrossRef] [PubMed]
- 19. Dizaj, S.M.; Barzegar-Jalali, M.; Zarrintan, M.H.; Adibkia, K.; Lotfipour, F. Calcium carbonate nanoparticles as cancer drug delivery system. *Expert Opin. Drug Deliv.* **2015**, *12*, 1649–1660. [CrossRef] [PubMed]
- 20. Hua, K.H.; Wang, H.C.; Chung, R.S.; Hsu, J.C. Calcium carbonate nanoparticles can enhance plant nutrition and insect pest tolerance. *J. Pestic. Sci.* 2015, *40*, 208–213. [CrossRef]
- 21. Setiawan, H.; Khairani, R.; Rahman, M.A.; Septawendar, R.; Mukti, R.R.; Dipojono, H.K.; Purwasasmita, B.S. Synthesis of zeolite and gamma-alumina nanoparticles as ceramic membranes for desalination applications. *J. Aust. Ceram. Soc.* **2017**, *53*, 531–538. [CrossRef]
- Singh, I.B.; Gupta, A.; Dubey, S.; Shafeeq, M.; Banerjee, P.; Sinha, A.S.K. Sol-gel synthesis of nanoparticles of gamma alumina and their application in defluoridation of water. *J. Sol-Gel Sci. Technol.* 2016, 77, 416–422. [CrossRef]
- 23. Nazari, A.; Sanjayan, J.G. Hybrid effects of alumina and silica nanoparticles on water absorption of geopolymers: Application of Taguchi approach. *Measurement* **2015**, *60*, 240–246. [CrossRef]
- 24. Palaniraja, J.; Arunachalam, P.; Vijayalakshmi, U.; Ghanem, M.A.; Roopan, S.M. Synthesis of calcium silicate nanoparticles and its catalytic application in Friedlander reaction. *Inorg. Nano-Met. Chem.* **2017**, *47*, 946–949. [CrossRef]
- Wu, J.; Zhu, Y.J.; Chen, F.; Zhao, X.Y.; Zhao, J.; Qi, C. Amorphous calcium silicate hydrate/block copolymer hybrid nanoparticles: Synthesis and application as drug carriers. *Dalton Trans.* 2013, 42, 7032–7040. [CrossRef] [PubMed]
- Strambeanu, N.; Demetrovici, L.; Dragos, D. Natural sources of nanoparticles. In Nanoparticles' Promises and Risks: Characterization, Manipulation, and Potential Hazards to Humanity and the Environment; Lungu, M., Neculae, A., Bunoiu, M., Biris, C., Eds.; Springer International Publishing: Cham, Switzerland, 2015; pp. 9–19.
- 27. Sun, R.; Yin, L.; Zhang, S.H.; He, L.; Cheng, X.J.; Wang, A.N.; Xia, H.W.; Shi, H.B. Simple light-triggered fluorescent labeling of silica nanoparticles for cellular imaging applications. *Chem. Eur. J.* **2017**, *23*, 13893–13896. [CrossRef] [PubMed]
- Xu, Z.G.; Ma, X.Q.; Gao, Y.E.; Hou, M.L.; Xue, P.; Li, C.M.; Kang, Y.J. Multifunctional silica nanoparticles as a promising theranostic platform for biomedical applications. *Mater. Chem. Front.* 2017, 1, 1257–1272. [CrossRef]
- 29. Liu, X.N.; Lu, X.R.; Wen, P.; Shu, X.Y.; Chi, F.T. Synthesis of ultrasmall silica nanoparticles for application as deep-ultraviolet antireflection coatings. *Appl. Surf. Sci.* **2017**, *420*, 180–185. [CrossRef]
- 30. Bergin, I.L.; Witzmann, F.A. Nanoparticle toxicity by the gastrointestinal route: Evidence and knowledge gaps. *Int. J. Biomed. Nanosci. Nanotechnol.* **2013**, *3*, 163–210. [CrossRef] [PubMed]
- Stawski, T.M.; Van Driessche, A.E.S.; Ossorio, M.; Rodriguez-Blanco, J.D.; Besselink, R.; Benning, L.G. Formation of calcium sulfate through the aggregation of sub-3 nanometre primary species. *Nat. Commun.* 2016, 7. [CrossRef] [PubMed]
- 32. Park, Y.B.; Mohan, K.; Al-Sanousi, A.; Almaghrabi, B.; Genco, R.J.; Swihart, M.T.; Dziak, R. Synthesis and characterization of nanocrystalline calcium sulfate for use in osseous regeneration. *Biomed. Mater.* **2011**, *6*. [CrossRef] [PubMed]
- 33. Guo, H.B.; Barnard, A.S. Naturally occurring iron oxide nanoparticles: Morphology, surface chemistry and environmental stability. *J. Mater. Chem. A* 2013, *1*, 27–42. [CrossRef]
- Groult, H.; Poupard, N.; Herranz, F.; Conforto, E.; Bridiau, N.; Sannier, F.; Bordenave, S.; Piot, J.M.; Ruiz-Cabello, J.; Fruitier-Arnaudin, I.; et al. Family of bioactive heparin-coated iron oxide nanoparticles with positive contrast in magnetic resonance imaging for specific biomedical applications. *Biomacromolecules* 2017, 18, 3156–3167. [CrossRef] [PubMed]
- 35. Saeedi, M.; Vahidi, O.; Bonakdar, S. Synthesis and characterization of glycyrrhizic acid coated iron oxide nanoparticles for hyperthermia applications. *Mater. Sci. Eng. C* 2017, 77, 1060–1067. [CrossRef] [PubMed]
- 36. Elrouby, M.; Abdel-Mawgoud, A.M.; Abd El-Rahman, R. Synthesis of iron oxides nanoparticles with very high saturation magnetization form TEA-Fe(III) complex via electrochemical deposition for supercapacitor applications. *J. Mol. Struct.* **2017**, *1147*, 84–95. [CrossRef]
- 37. Perez, J.M. Iron oxide nanoparticles—Hidden talent. Nat. Nanotechnol. 2007, 2, 535–536. [CrossRef] [PubMed]
- 38. Wikipedia. Umber. Available online: https://en.wikipedia.org/wiki/Umber (accessed on 19 December 2017).

- 39. Yuan, J.K.; Yang, J.; Suib, S.L. Synthesis of microporous manganese oxide nanoparticles and their catalysis applications. *Abstr. Pap. Am. Chem. Soc.* **2004**, 227, U1310.
- Zhao, D.Y.; Han, B.; Xie, W.B.; An, B. Applications of stabilized manganese oxide and Fe-Mn binary oxides nanoparticles for in situ remediation of contaminated soil and groundwater. *Abstr. Pap. Am. Chem. Soc.* 2014, 247. [CrossRef]
- Luo, Y.; Yang, J.; Li, J.C.; Yu, Z.B.; Zhang, G.X.; Shi, X.Y.; Shen, M.W. Facile synthesis and functionalization of manganese oxide nanoparticles for targeted T₁-weighted tumor MR imaging. *Colloid Surf. B* 2015, 136, 506–513. [CrossRef] [PubMed]
- 42. Faulstich, L.; Griffin, S.; Nasim, M.J.; Masood, M.I.; Ali, W.; Alhamound, S.; Omran, Y.; Kim, H.; Kharma, A.; Schafer, K.H.; et al. Nature's hat-trick: Can we use sulfur springs as ecological source for materials with agricultural and medical applications? *Int. Biodeterior. Biodegrad.* **2017**, *119*, 678–686. [CrossRef]
- 43. Suleiman, M.; Al Ali, A.; Hussein, A.; Hammouti, B.; Hadda, T.B.; Warad, I. Sulfur nanoparticles: Synthesis, characterizations and their applications. *J. Mater. Environ. Sci.* **2013**, *4*, 1029–1033.
- 44. Terrones, M. Carbon nanotubes: Synthesis and properties, electronic devices and other emerging applications. *Int. Mater. Rev.* **2004**, *49*, 325–377. [CrossRef]
- 45. Popov, V.N. Carbon nanotubes: Properties and application. Mater. Sci. Eng. R 2004, 43, 61–102. [CrossRef]
- 46. McGillicuddy, E.; Murray, I.; Kavanagh, S.; Morrison, L.; Fogarty, A.; Cormican, M.; Dockery, P.; Prendergast, M.; Rowan, N.; Morris, D. Silver nanoparticles in the environment: Sources, detection and ecotoxicology. *Sci. Total Environ.* **2017**, *575*, 231–246. [CrossRef] [PubMed]
- 47. Zhang, C.Q.; Hu, Z.Q.; Deng, B.L. Silver nanoparticles in aquatic environments: Physiochemical behavior and antimicrobial mechanisms. *Water Res.* **2016**, *88*, 403–427. [CrossRef] [PubMed]
- 48. Rai, M.; Ingle, A.P.; Paralikar, P. Sulfur and sulfur nanoparticles as potential antimicrobials: From traditional medicine to nanomedicine. *Expert Rev. Anti-Infect.* **2016**, *14*, 969–978. [CrossRef] [PubMed]
- 49. Hough, R.M.; Noble, R.R.P.; Reich, M. Natural gold nanoparticles. Ore Geol. Rev. 2011, 42, 55–61. [CrossRef]
- 50. Dykman, L.A.; Khlebtsov, N.G. Gold nanoparticles in biology and medicine: Recent advances and prospects. *Acta Nat.* **2011**, *3*, 34–55.
- 51. Rauch, S.; Hemond, H.F.; Barbante, C.; Owari, M.; Morrison, G.M.; Peucker-Ehrenbrink, B.; Wass, U. Importance of automobile exhaust catalyst emissions for the deposition of platinum, palladium, and rhodium in the northern hemisphere. *Environ. Sci. Technol.* **2005**, *39*, 8156–8162. [CrossRef] [PubMed]
- 52. Cheng, Q.; Liu, Y. Multifunctional platinum-based nanoparticles for biomedical applications. *Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.* **2017**, *9*. [CrossRef] [PubMed]
- Sheny, D.S.; Philip, D.; Mathew, J. Synthesis of platinum nanoparticles using dried anacardium occidentale leaf and its catalytic and thermal applications. *Spectrochim. Acta Part A* 2013, 114, 267–271. [CrossRef] [PubMed]
- 54. Pedone, D.; Moglianetti, M.; De Luca, E.; Bardi, G.; Pompa, P.P. Platinum nanoparticles in nanobiomedicine. *Chem. Soc. Rev.* **2017**, *46*, 4951–4975. [CrossRef] [PubMed]
- 55. Blanco-Andujar, C.; Ortega, D.; Pankhurst, Q.A.; Thanh, N.T.K. Elucidating the morphological and structural evolution of iron oxide nanoparticles formed by sodium carbonate in aqueous medium. *J. Mater. Chem.* **2012**, 22, 12498–12506. [CrossRef]
- 56. Cho, M.H.; Choi, E.-S.; Kim, S.; Goh, S.-H.; Choi, Y. Redox-responsive manganese dioxide nanoparticles for enhanced MR imaging and radiotherapy of lung cancer. *Front. Chem.* **2017**, *5*. [CrossRef] [PubMed]
- 57. Song, S.Q.; Rao, R.C.; Yang, H.X.; Liu, H.D.; Zhang, A.M. Facile synthesis of Fe₃O₄/MWCNTs by spontaneous redox and their catalytic performance. *Nanotechnology* **2010**, *21*. [CrossRef] [PubMed]
- 58. Santillo, D.; Miller, K.; Johnston, P. Microplastics as contaminants in commercially important seafood species. *Integr. Environ. Assess.* **2017**, *13*, 516–521. [CrossRef] [PubMed]
- 59. Cole, M.; Lindeque, P.; Halsband, C.; Galloway, T.S. Microplastics as contaminants in the marine environment: A review. *Mar. Pollut. Bull.* **2011**, *62*, 2588–2597. [CrossRef] [PubMed]
- Pawlak, J.; Lodyga-Chrucinska, E.; Chrustowicz, J. Fate of platinum metals in the environment. J. Trace Elem. Med. Biol. 2014, 28, 247–254. [CrossRef] [PubMed]
- 61. Zimmermann, S.; Sures, B. Significance of platinum group metals emitted from automobile exhaust gas converters for the biosphere. *Environ. Sci. Pollut. Res.* 2004, *11*, 194–199. [CrossRef]
- 62. Ezoe, Y.; Lin, C.H.; Noto, M.; Watanabe, Y.; Yoshimura, K. Evolution of water chemistry in natural acidic environments in Yangmingshan, Taiwan. *J. Environ. Monit.* **2002**, *4*, 533–540. [CrossRef] [PubMed]

- 63. Berlo, K.; van Hinsberg, V.J.; Vigouroux, N.; Gagnon, J.E.; Williams-Jones, A.E. Sulfide breakdown controls metal signature in volcanic gas at Kawah Ijen volcano, Indonesia. *Chem. Geol.* **2014**, *371*, 115–127. [CrossRef]
- 64. Mitchell, S.C.; Waring, R.H. Sulphate absorption across biological membranes. *Xenobiotica* **2016**, *46*, 184–191. [CrossRef] [PubMed]
- 65. Jacob, C. Redox signalling via the cellular thiolstat. *Biochem. Soc. Trans.* **2011**, *39*, 1247–1253. [CrossRef] [PubMed]
- 66. Giles, G.; Nasim, M.; Ali, W.; Jacob, C. The reactive sulfur species concept: 15 years on. *Antioxidants* **2017**, *6*. [CrossRef] [PubMed]
- 67. Cummins, L.M.; Kimura, E.T. Safety evaluation of selenium sulfide antidandruff shampoos. *Toxicol. Appl. Pharmacol.* **1971**, *20*, 89–96. [CrossRef]
- 68. Evolution. Available online: https://en.Wikipedia.Org/wiki/evolution_(2001_film) (accessed on 20 December 2017).
- Schneider, T.; Baldauf, A.; Ba, L.A.; Jamier, V.; Khairan, K.; Sarakbi, M.B.; Reum, N.; Schneider, M.; Roseler, A.; Becker, K.; et al. Selective antimicrobial activity associated with sulfur nanoparticles. *J. Biomed. Nanotechnol.* 2011, 7, 395–405. [CrossRef] [PubMed]
- 70. Brown, T. The human genome. In Genomes, 2nd ed.; Wiley: Oxford, UK, 2002.
- 71. Shors, T. Virus architecture and nomenclature. In *Understanding Viruses*, 2nd ed.; Jones & Bartlett Learning: Burlington, MA, USA, 2011.
- 72. Janssen, A.; de Keizer, A.; van Aelst, A.; Fokkink, R.; Yangling, H.; Lettinga, G. Surface characteristics and aggregation of microbiologically produced sulphur particles in relation to the process conditions. *Colloid Surf. B* **1996**, *6*, 115–129. [CrossRef]
- Mishra, S.; Singh, B.R.; Naqvi, A.H.; Singh, H.B. Potential of biosynthesized silver nanoparticles using *Stenotrophomonas* sp. BHU-S7 (MTCC 5978) for management of soil-borne and foliar phytopathogens. *Sci. Rep.* 2017, 7. [CrossRef] [PubMed]
- 74. Mishra, S.; Singh, B.R.; Singh, A.; Keswani, C.; Naqvi, A.H.; Singh, H.B. Biofabricated silver nanoparticles act as a strong fungicide against *Bipolaris sorokiniana* causing spot blotch disease in wheat. *PLoS ONE* **2014**, *9*, e97881. [CrossRef] [PubMed]
- Malhotra, A.; Dolma, K.; Kaur, N.; Rathore, Y.S.; Ashish; Mayilraj, S.; Choudhury, A.R. Biosynthesis of gold and silver nanoparticles using a novel marine strain of *Stenotrophomonas*. *Bioresour. Technol.* 2013, 142, 727–731. [CrossRef] [PubMed]
- Singh, P.K.; Kundu, S. Biosynthesis of gold nanoparticles using bacteria. *Proc. Natl. Acad. Sci. India Sect. B* 2014, 84, 331–336. [CrossRef]
- 77. Stefess, G.C.; Torremans, R.A.M.; DeSchrijver, R.; Robertson, L.A.; Kuenen, J.G. Quantitative measurement of sulphur formation by steady state and transient state continuous cultures of autotrophic thiobacillus species. *Appl. Microbiol. Biot.* **1996**, 45, 169–175. [CrossRef]
- Kuppusamy, P.; Yusoff, M.M.; Maniam, G.P.; Govindan, N. Biosynthesis of metallic nanoparticles using plant derivatives and their new avenues in pharmacological applications—An updated report. *Saudi Pharm. J.* 2016, 24, 473–484. [CrossRef] [PubMed]
- 79. Wright, M.H.; Farooqui, S.M.; White, A.R.; Greene, A.C. Production of manganese oxide nanoparticles by *Shewanella* species. *Appl. Environ. Microbiol.* **2016**, *82*, 5402–5409. [CrossRef] [PubMed]
- Garmasheva, I.; Kovalenko, N.; Voychuk, S.; Ostapchuk, A.; Livins'ka, O.; Oleschenko, L. Lactobacillus species mediated synthesis of silver nanoparticles and their antibacterial activity against opportunistic pathogens in vitro. *Bioimpacts* 2016, *6*, 219–223. [CrossRef] [PubMed]
- 81. Singh, B.K.; Walker, A. Microbial degradation of organophosphorus compounds. *FEMS Microbiol. Rev.* **2006**, 30, 428–471. [CrossRef] [PubMed]
- 82. Yadav, K.K.; Singh, J.K.; Gupta, N.; Kumar, V. A review of nanobioremediation technologies for environmental cleanup: A novel biological approach. *J. Mater. Environ. Sci.* **2017**, *8*, 740–757.
- 83. Sanghi, R.; Verma, P.; Puri, S. Enzymatic formation of gold nanoparticles using phanerochaete chrysosporium. *Adv. Chem. Eng. Sci.* **2011**, *1*, 1–8. [CrossRef]
- 84. Durán, N.; Marcato, P.D.; Alves, O.L.; De Souza, G.I.H.; Esposito, E. Mechanistic aspects of biosynthesis of silver nanoparticles by several *Fusarium oxysporum* strains. *J. Nanobiotechnol.* **2005**, *3*. [CrossRef] [PubMed]
- 85. Duhan, J.S.; Kumar, R.; Kumar, N.; Kaur, P.; Nehra, K.; Duhan, S. Nanotechnology: The new perspective in precision agriculture. *Biotechnol. Rep.* **2017**, *15*, 11–23. [CrossRef] [PubMed]

- Iavicoli, I.; Leso, V.; Beezhold, D.H.; Shvedova, A.A. Nanotechnology in agriculture: Opportunities, toxicological implications, and occupational risks. *Toxicol. Appl. Pharmacol.* 2017, 329, 96–111. [CrossRef] [PubMed]
- 87. Salata, O. Applications of nanoparticles in biology and medicine. *J. Nanobiotechnol.* **2004**, 2. [CrossRef] [PubMed]
- 88. Li, X.Q.; Xu, H.Z.; Chen, Z.S.; Chen, G.F. Biosynthesis of nanoparticles by microorganisms and their applications. *J. Nanomater.* **2011**. [CrossRef]
- Estevam, E.C.; Griffin, S.; Nasim, M.J.; Denezhkin, P.; Schneider, R.; Lilischkis, R.; Dominguez-Alvarez, E.; Witek, K.; Latacz, G.; Keck, C.; et al. Natural selenium particles from *Staphylococcus carnosus*: Hazards or particles with particular promise? *J. Hazard. Mater.* 2017, *324*, 22–30. [CrossRef] [PubMed]
- Zhang, L.; Li, D.P.; Gao, P. Expulsion of selenium/protein nanoparticles through vesicle-like structures by saccharomyces cerevisiae under microaerophilic environment. *World J. Microbiol. Biotechnol.* 2012, 28, 3381–3386. [CrossRef] [PubMed]
- 91. Skalickova, S.; Milosavljevic, V.; Cihalova, K.; Horky, P.; Richtera, L.; Adam, V. Selenium nanoparticles as a nutritional supplement. *Nutrition* **2017**, *33*, 83–90. [CrossRef] [PubMed]
- 92. Wang, T.T.; Yang, L.B.; Zhang, B.C.; Liu, J.H. Extracellular biosynthesis and transformation of selenium nanoparticles and application in H₂O₂ biosensor. *Colloid Surf. B* **2010**, *80*, 94–102. [CrossRef] [PubMed]
- Yazdi, M.H.; Mahdavi, M.; Varastehmoradi, B.; Faramarzi, M.A.; Shahverdi, A.R. The immunostimulatory effect of biogenic selenium nanoparticles on the 4T1 breast cancer model: An in vivo study. *Biol. Trace Elem. Res.* 2012, 149, 22–28. [CrossRef] [PubMed]
- Prakash, N.T.; Sharma, N.; Prakash, R.; Raina, K.K.; Fellowes, J.; Pearce, C.I.; Lloyd, J.R.; Pattrick, R.A.D. Aerobic microbial manufacture of nanoscale selenium: Exploiting nature's bio-nanomineralization potential. *Biotechnol. Lett.* 2009, *31*, 1857–1862. [CrossRef] [PubMed]
- Manikova, D.; Letavayova, L.M.; Vlasakova, D.; Kosik, P.; Estevam, E.C.; Nasim, M.J.; Gruhlke, M.; Slusarenko, A.; Burkholz, T.; Jacob, C.; et al. Intracellular diagnostics: Hunting for the mode of action of redox-modulating selenium compounds in selected model systems. *Molecules* 2014, 19, 12258–12279. [CrossRef] [PubMed]
- Castellucci Estevam, E.; Witek, K.; Faulstich, L.; Nasim, M.J.; Latacz, G.; Dominguez-Alvarez, E.; Kiec-Kononowicz, K.; Demasi, M.; Handzlik, J.; Jacob, C. Aspects of a distinct cytotoxicity of selenium salts and organic selenides in living cells with possible implications for drug design. *Molecules* 2015, 20, 13894–13912. [CrossRef] [PubMed]
- 97. Reddy, C.A.; Mathew, Z. Bioremediation potential of white rot fungi. In *British Mycological Society Symposium Series*; Cambridge University Press: Cambridge, UK, 2001; pp. 52–78.
- Padhi, S.K.; Tripathy, S.; Sen, R.; Mahapatra, A.S.; Mohanty, S.; Maiti, N.K. Characterisation of heterotrophic nitrifying and aerobic denitrifying *Klebsiella pneumoniae* CF-S9 strain for bioremediation of wastewater. *Int. Biodeterior. Biodegrad.* 2013, 78, 67–73. [CrossRef]
- Duran, N.; Cuevas, R.; Cordi, L.; Rubilar, O.; Diez, M.C. Biogenic silver nanoparticles associated with silver chloride nanoparticles (Ag@AgCl) produced by laccase from *Trametes versicolor*. *SpringerPlus* 2014, 3. [CrossRef] [PubMed]
- 100. Anil Kumar, S.; Abyaneh, M.K.; Gosavi, S.W.; Kulkarni, S.K.; Pasricha, R.; Ahmad, A.; Khan, M.I. Nitrate reductase-mediated synthesis of silver nanoparticles from AgNO₃. *Biotechnol. Lett.* 2007, 29, 439–445. [CrossRef] [PubMed]
- 101. Makarov, V.V.; Love, A.J.; Sinitsyna, O.V.; Makarova, S.S.; Yaminsky, I.V.; Taliansky, M.E.; Kalinina, N.O. "Green" nanotechnologies: Synthesis of metal nanoparticles using plants. *Acta Nat.* **2014**, *6*, 35–44.
- 102. Khodashenas, B.; Ghorbani, H.R. Synthesis of silver nanoparticles with different shapes. *Arab. J. Chem.* **2015**. [CrossRef]
- 103. Singh, A.K.; Kanchanapally, R.; Fan, Z.; Senapati, D.; Ray, P.C. Synthesis of highly fluorescent water-soluble silver nanoparticles for selective detection of Pb(II) at the parts per quadrillion (PPQ) level. *Chem. Commun.* 2012, 48, 9047–9049. [CrossRef] [PubMed]
- 104. Zhou, T.; Rong, M.; Cai, Z.; Yang, C.J.; Chen, X. Sonochemical synthesis of highly fluorescent glutathione-stabilized Ag nanoclusters and S^{2–} sensing. *Nanoscale* **2012**, *4*, 4103–4106. [CrossRef] [PubMed]
- 105. Jain, S.; Mehata, M.S. Medicinal plant leaf extract and pure flavonoid mediated green synthesis of silver nanoparticles and their enhanced antibacterial property. *Sci. Rep.* **2017**, *7*. [CrossRef] [PubMed]

- 106. Si, S.; Mandal, T.K. Tryptophan-based peptides to synthesize gold and silver nanoparticles: A mechanistic and kinetic study. *Chem. Eur. J.* 2007, *13*, 3160–3168. [CrossRef] [PubMed]
- Brodin, J.D.; Carr, J.R.; Sontz, P.A.; Tezcan, F.A. Exceptionally stable, redox-active supramolecular protein assemblies with emergent properties. *Proc. Natl. Acad. Sci. USA* 2014, 111, 2897–2902. [CrossRef] [PubMed]
- 108. Lee, K.J.; Park, S.H.; Govarthanan, M.; Hwang, P.H.; Seo, Y.S.; Cho, M.; Lee, W.H.; Lee, J.Y.; Kamala-Kannan, S.; Oh, B.T. Synthesis of silver nanoparticles using cow milk and their antifungal activity against phytopathogens. *Mater. Lett.* **2013**, *105*, 128–131. [CrossRef]
- 109. Arrigoni, O.; De Tullio, M.C. Ascorbic acid: Much more than just an antioxidant. *Biochim. Biophys. Acta Gen. Subj.* **2002**, 1569, 1–9. [CrossRef]
- 110. Yamada, C.; Kawai, H.; Yoshida, K. Improving ascorbic acid content of tomato fruits by the oxidized yeast extract. In *Plant Nutrition for Sustainable Food Production and Environment, Proceedings of the XIII International Plant Nutrition Colloquium, Tokyo, Japan, 13–19 September 1997*; Ando, T., Fujita, K., Mae, T., Matsumoto, H., Mori, S., Sekiya, J., Eds.; Springer: Dordrecht, The Netherlands, 1997; pp. 973–974.
- 111. Balcerczyk, A.; Grzelak, A.; Janaszewska, A.; Jakubowski, W.; Koziol, S.; Marszalek, M.; Rychlik, B.; Soszynski, M.; Bilinski, T.; Bartosz, G. Thiols as major determinants of the total antioxidant capacity (reprinted from thiol metabolism and redox regulation of cellular functions). *Biofactors* 2003, *17*, 75–82. [CrossRef] [PubMed]
- 112. Swiegers, J.H.; Capone, D.L.; Pardon, K.H.; Elsey, G.M.; Sefton, M.A.; Francis, I.L.; Pretorius, I.S. Engineering volatile thiol release in *Saccharomyces cerevisiae* for improved wine aroma. *Yeast* 2007, 24, 561–574. [CrossRef] [PubMed]
- 113. Kerr, E.D.; Schulz, B.L. Vegemite beer: Yeast extract spreads as nutrient supplements to promote fermentation. *PeerJ* **2016**, *4*, e2271. [CrossRef] [PubMed]
- 114. Shulman, K.I.; Walker, S.E.; Mackenzie, S.; Knowles, S. Dietary restriction, tyramine, and the use of monoamine-oxidase inhibitors. *J. Clin. Psychopharmacol.* **1989**, *9*, 397–402. [CrossRef] [PubMed]
- 115. Musarrat, J.; Dwivedi, S.; Singh, B.R.; Al-Khedhairy, A.A.; Azam, A.; Naqvi, A. Production of antimicrobial silver nanoparticles in water extracts of the fungus *Amylomyces rouxii* strain KSU-09. *Bioresour. Technol.* 2010, 101, 8772–8776. [CrossRef] [PubMed]
- Sreekanth, T.V.; Ravikumar, S.; Eom, I.Y. Green synthesized silver nanoparticles using nelumbonucifera root extract for efficient protein binding, antioxidant and cytotoxicity activities. *J. Photochem. Photobiol. B* 2014, 141, 100–105. [CrossRef] [PubMed]
- Ravikumar, S.; Sreekanth, T.V.M.; Eom, I.Y. Interaction studies of greenly synthesized gold nanoparticles with bovine serum albumin (BSA) using fluorescence spectroscopy. *J. Nanosci. Nanotechnol.* 2015, 15, 9617–9623. [CrossRef] [PubMed]
- Dhayalan, M.; Denison, M.I.J.; Jegadeeshwari, L.A.; Krishnan, K.; Gandhi, N.N. In vitro antioxidant, antimicrobial, cytotoxic potential of gold and silver nanoparticles prepared using *Embelia ribes*. *Nat. Prod. Res.* 2017, 31, 465–468. [CrossRef] [PubMed]
- Stan, M.; Popa, A.; Toloman, D.; Silipas, T.D.; Vodnar, D.C. Antibacterial and antioxidant activities of ZnO nanoparticles synthesized using extracts of *Allium Sativum*, *Rosmarinus officinalis* and *Ocimum basilicum*. *Acta Metall Sin. Engl.* 2016, 29, 228–236. [CrossRef]
- 120. Stan, M.; Popa, A.; Toloman, D.; Silipas, T.-D.; Vodnar, D.C.; Katona, G. Enhanced antibacterial activity of zinc oxide nanoparticles synthesized using *Petroselinum crispum* extracts. In *AIP Conference Proceedings*; AIP Publishing: College Park, MD, USA, 2015; Volume 1700.
- 121. Stan, M.; Lung, I.; Soran, M.L.; Leostean, C.; Popa, A.; Stefan, M.; Lazar, M.D.; Opris, O.; Silipas, T.D.; Porav, A.S. Removal of antibiotics from aqueous solutions by green synthesized magnetite nanoparticles with selected agro-waste extracts. *Process Saf. Environ.* 2017, 107, 357–372. [CrossRef]
- 122. Prakash, P.; Gnanaprakasam, P.; Emmanuel, R.; Arokiyaraj, S.; Saravanan, M. Green synthesis of silver nanoparticles from leaf extract of *Mimusops elengi*, Linn. for enhanced antibacterial activity against multi drug resistant clinical isolates. *Colloid Surf. B* **2013**, *108*, 255–259. [CrossRef] [PubMed]
- 123. Krishnaraj, C.; Ramachandran, R.; Mohan, K.; Kalaichelvan, P.T. Optimization for rapid synthesis of silver nanoparticles and its effect on phytopathogenic fungi. *Spectrochim. Acta Part A* 2012, 93, 95–99. [CrossRef] [PubMed]

- 124. Otunola, G.A.; Afolayan, A.J.; Ajayi, E.O.; Odeyemi, S.W. Characterization, antibacterial and antioxidant properties of silver nanoparticles synthesized from aqueous extracts of *Allium sativum*, *Zingiber officinale*, and *Capsicum frutescens*. *Pharmacogn*. *Mag.* 2017, *13*, S201–S208. [CrossRef] [PubMed]
- 125. Barbulova, A.; Colucci, G.; Apone, F. New trends in cosmetics: By-products of plant origin and their potential use as cosmetic active ingredients. *Cosmetics* **2015**, *2*, 82–92. [CrossRef]
- 126. Chokshi, K.; Pancha, I.; Ghosh, T.; Paliwal, C.; Maurya, R.; Ghosh, A.; Mishra, S. Green synthesis, characterization and antioxidant potential of silver nanoparticles biosynthesized from de-oiled biomass of thermotolerant oleaginous microalgae acutodesmus dimorphus. *RSC Adv.* **2016**, *6*, 72269–72274. [CrossRef]
- 127. Harish, S.P. Phyco-nanotechnology: New horizons of gold nano-factories. *Proc. Natl. Acad. Sci. India* Sect. B 2016. [CrossRef]
- 128. Shankar, S.S.; Rai, A.; Ahmad, A.; Sastry, M. Rapid synthesis of Au, Ag, and bimetallic Au core-Ag shell nanoparticles using neem (*Azadirachta indica*) leaf broth. J. Colloid Interface Sci. 2004, 275, 496–502. [CrossRef] [PubMed]
- 129. Nune, S.K.; Chanda, N.; Shukla, R.; Katti, K.; Kulkarni, R.R.; Thilakavathy, S.; Mekapothula, S.; Kannan, R.; Katti, K.V. Green nanotechnology from tea: Phytochemicals in tea as building blocks for production of biocompatible gold nanoparticles. *J. Mater. Chem* 2009, *19*, 2912–2920. [CrossRef] [PubMed]
- Griffin, S.; Tittikpina, N.K.; Al-Marby, A.; Alkhayer, R.; Denezhkin, P.; Witek, K.; Gbogbo, K.A.; Batawila, K.; Duval, R.E.; Nasim, M.J.; et al. Turning waste into value: Nanosized natural plant materials of *Solanum incanum* L. and *Pterocarpus erinaceus* poir with promising antimicrobial activities. *Pharmaceutics* 2016, 8. [CrossRef] [PubMed]
- Mauludin, R.; Müller, R.H.; Keck, C.M. Development of an oral rutin nanocrystal formulation. *Int. J. Pharm.* 2009, 370, 202–209. [CrossRef] [PubMed]
- 132. Mueller, R.H.; Keck, C.M. Second generation of drug nanocrystals for delivery of poorly soluble drugs: smartCrystal technology. *Eur. J. Pharm. Sci.* 2008, 34, S20–S21. [CrossRef]
- 133. Keck, C.M.; Muller, R.H. Drug nanocrystals of poorly soluble drugs produced by high pressure homogenisation. *Eur. J. Pharm. Biopharm.* **2006**, *62*, 3–16. [CrossRef] [PubMed]
- 134. Muller, R.H.; Gohla, S.; Keck, C.M. State of the art of nanocrystals—Special features, production, nanotoxicology aspects and intracellular delivery. *Eur. J. Pharm. Biopharm.* **2011**, *78*, 1–9. [CrossRef] [PubMed]
- 135. Muller, R.H.; Keck, C.M. Challenges and solutions for the delivery of biotech drugs—A review of drug nanocrystal technology and lipid nanoparticles. *J. Biotechnol.* **2004**, *113*, 151–170. [CrossRef] [PubMed]
- 136. Muller, R.H.; Keck, C.M. Twenty years of drug nanocrystals: Where are we, and where do we go? *Eur. J. Pharm. Biopharm.* **2012**, *80*, 1–3. [CrossRef] [PubMed]
- 137. Scholz, P.; Keck, C.M. Nanocrystals: From raw material to the final formulated oral dosage form—A review. *Curr. Pharm. Des.* **2015**, *21*, 4217–4228. [CrossRef] [PubMed]
- 138. Griffin, S.; Alkhayer, R.; Mirzoyan, S.; Turabyan, A.; Zucca, P.; Sarfraz, M.; Nasim, M.; Trchounian, A.; Rescigno, A.; Keck, C.; et al. Nanosizing *Cynomorium*: Thumbs up for potential antifungal applications. *Inventions* 2017, 2. [CrossRef]
- 139. Abdelwahed, W.; Degobert, G.; Stainmesse, S.; Fessi, H. Freeze-drying of nanoparticles: Formulation, process and storage considerations. *Adv. Drug Deliv. Rev.* **2006**, *58*, 1688–1713. [CrossRef] [PubMed]
- 140. Alihosseini, F.; Ghaffari, S.; Dabirsiaghi, A.R.; Haghighat, S. Freeze-drying of ampicillin solid lipid nanoparticles using mannitol as cryoprotectant. *Braz. J. Pharm. Sci.* **2015**, *51*, 797–802. [CrossRef]
- 141. Patrignani, F.; Lanciotti, R. Applications of high and ultra high pressure homogenization for food safety. *Front. Microbiol.* **2016**, *7*. [CrossRef] [PubMed]
- 142. Wikipedia. Milli Vanilli. Available online: https://en.wikipedia.org/wiki/Milli_Vanilli (accessed on 20 December 2017).
- 143. Ganesh Kumar, C.; Poornachandra, Y.; Mamidyala, S.K. Green synthesis of bacterial gold nanoparticles conjugated to resveratrol as delivery vehicles. *Colloids Surf. B* 2014, 123, 311–317. [CrossRef] [PubMed]



© 2017 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).